

## 3.1 Important traits of species of *Hippophae* genus

➤ Seeds of sea buckthorn are very small, with low oil content (7-10%), which leads to seed oil yield is less than 52.5kg per ha.

➤ The percentage of palmitic acid in pulp oil is up to 36%, which is not good for human health.

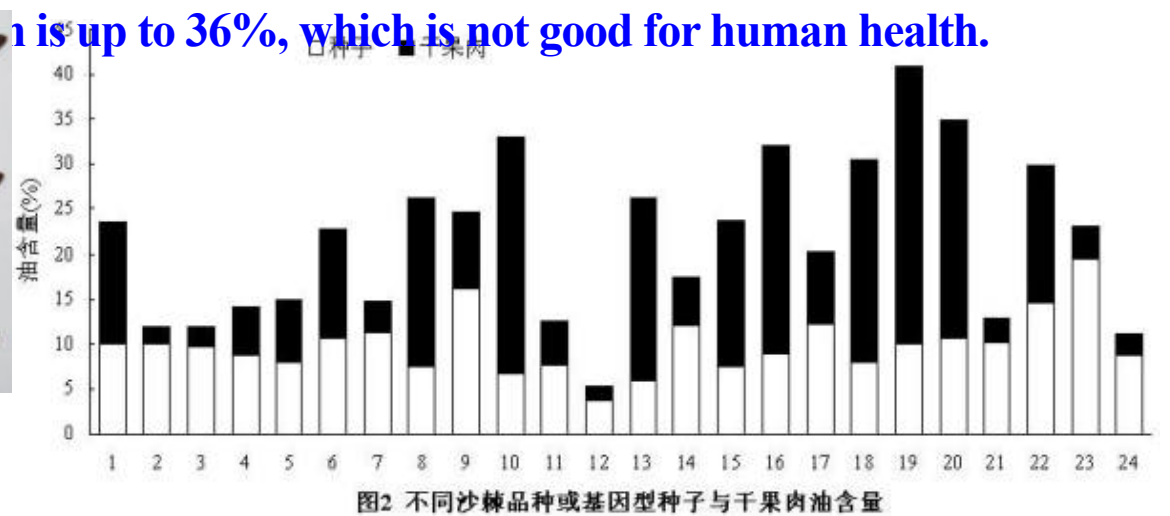
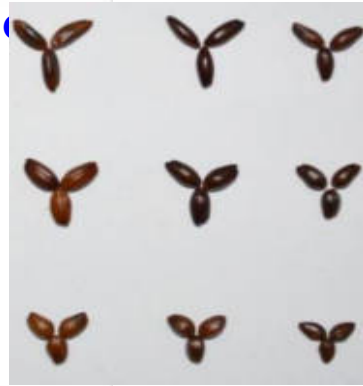
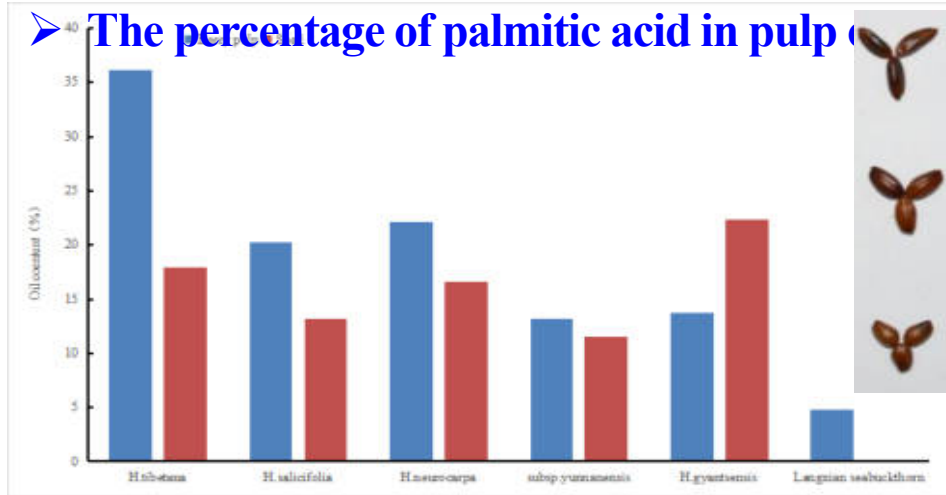
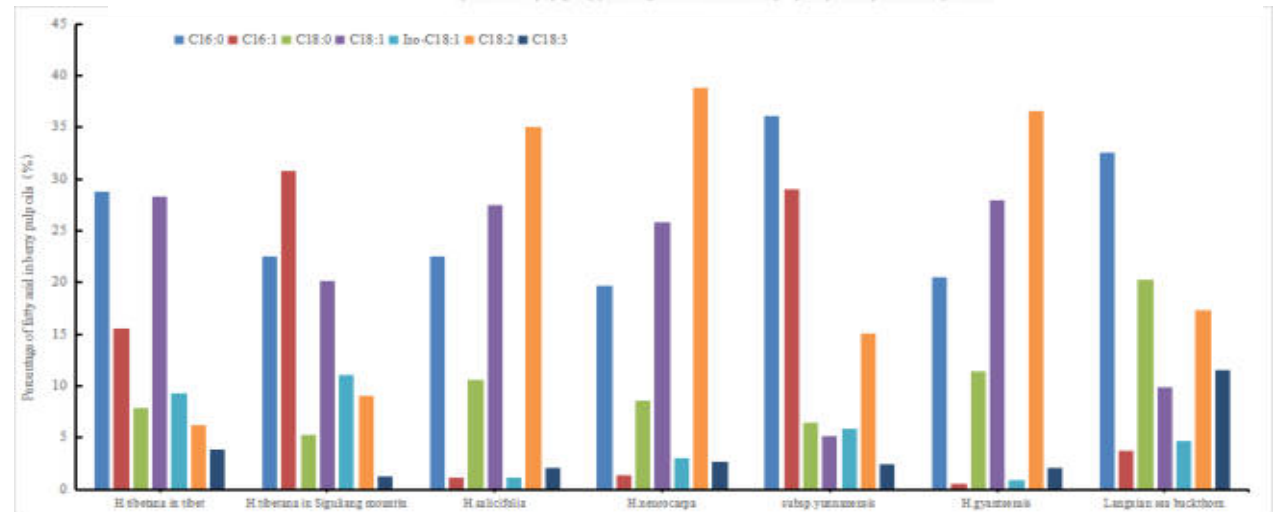
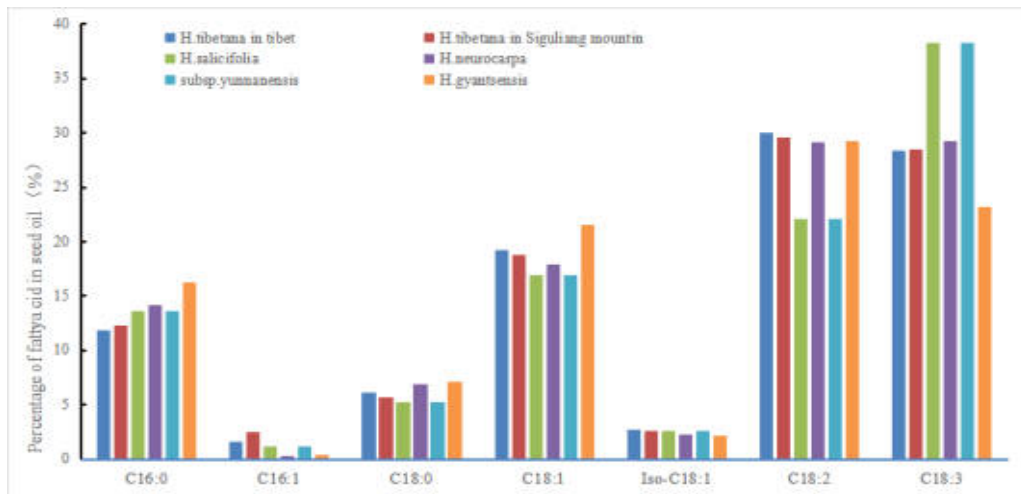


图2 不同沙棘品种或基因型种子与干果肉油含量



## 3.2 Co-regulation mechanism of source (*GPD1*) and pool (*DGAT*) genes for lipid biosynthesis in sea buckthorn seeds

High expression of *GPD1* gene occurred in the period of rapid lipid accumulation, which may speed up lipid biosynthesis in ‘Xin’e 3’ seeds through promoting the biosynthesis of glycerol-3-phosphate. The expression levels of *DGAT1* and *DGAT2* genes in line ‘Xin’e 3’ were higher than that line ‘Suiji 1’ during lipid accumulation.

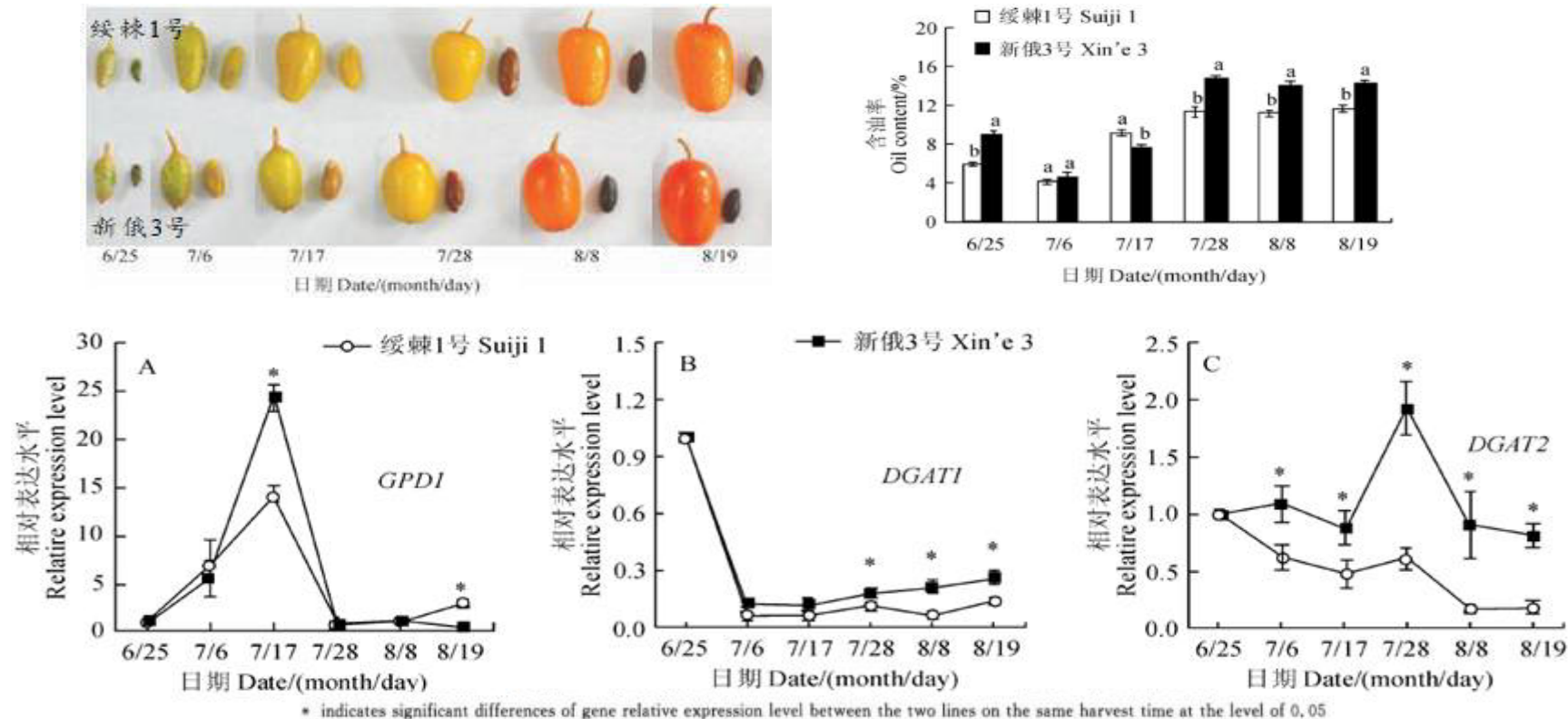


Fig. 3 Differences of *GPD1*, *DGAT1* and *DGAT2* genes expression in seeds between the line ‘Suiji 1’ and ‘Xin’e 3’



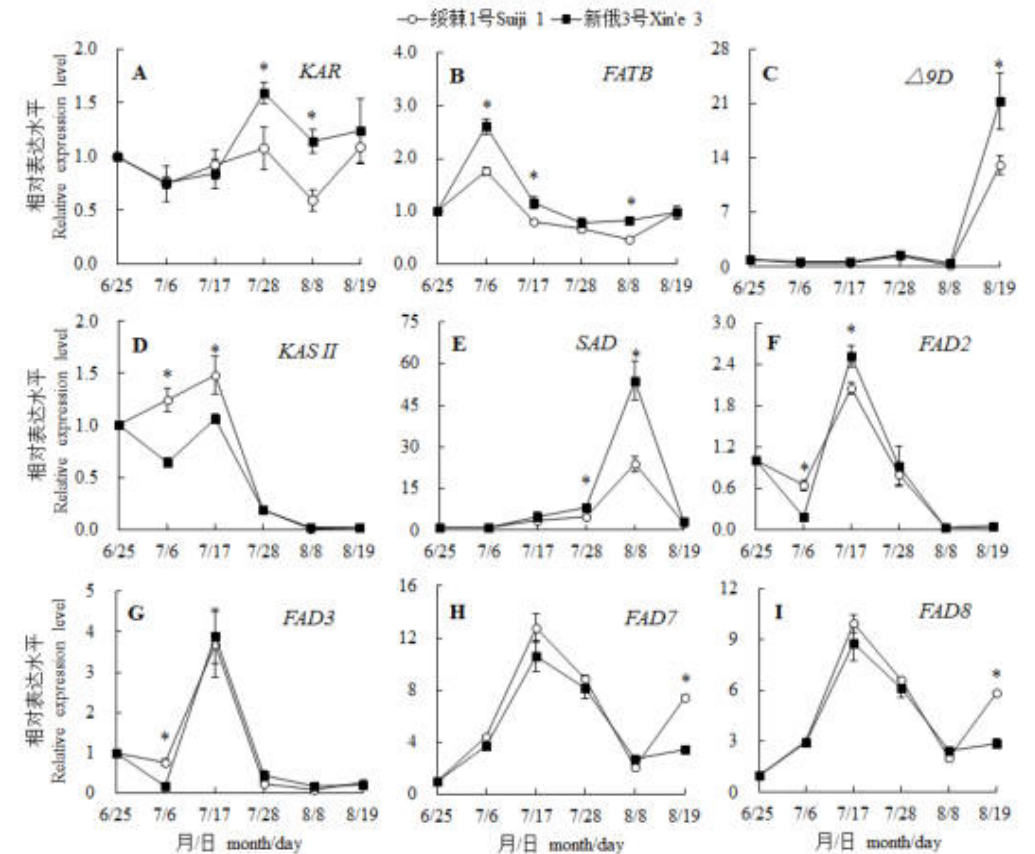
# 3. Formation mechanism of important traits in species of *Hippophae* genus



## 3.3 Multi gene coordinated regulation of high accumulation of C18 unsaturated fatty acids in sea buckthorn seeds

High accumulation of C18 unsaturated fatty acid in sea buckthorn seed oils originates from low expression of *FATB* and  $\Delta 9D$  genes and high co-expression of *KAR*, *KASII*, *SAD*, *FAD2*, *FAD3*, *FAD7* and *FAD8* genes.

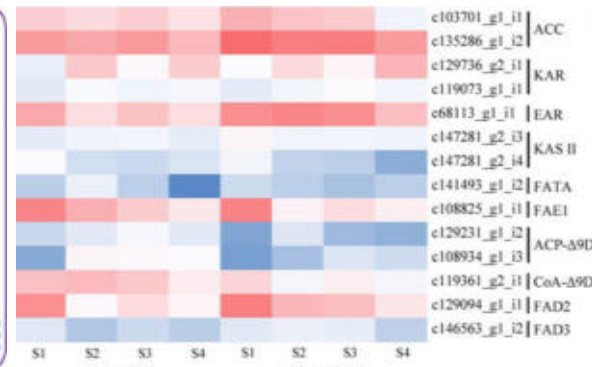
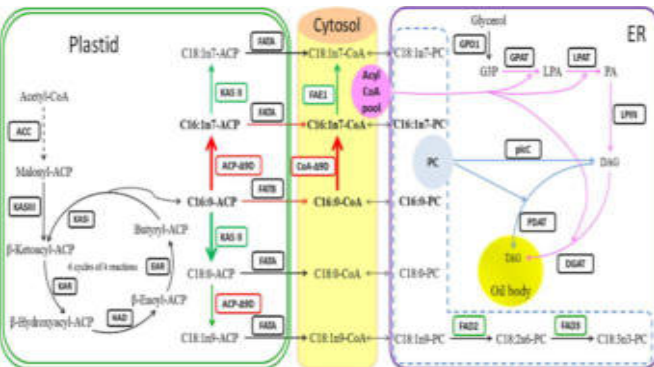
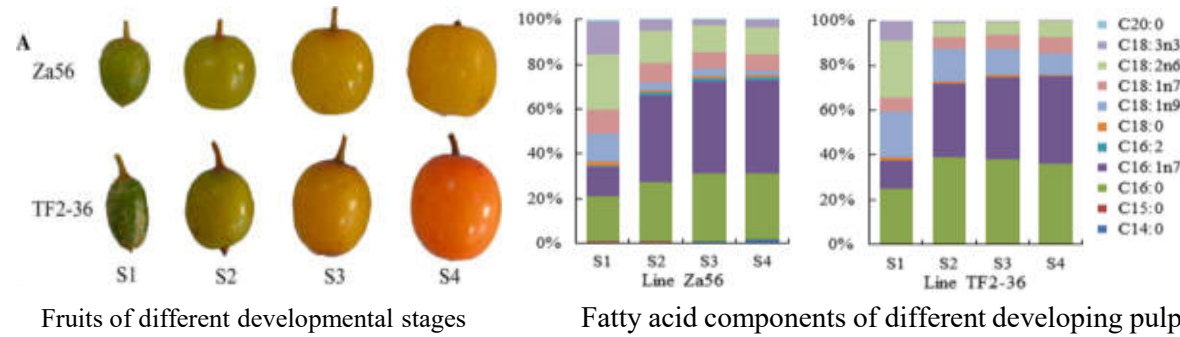
- The down-regulation expression of *FATB* coordinated with the low expression of  $\Delta 9D$  genes decreased the metabolic biosynthesis of palmitic and palmitoleic acids transformed from C16:0-ACP. Relatively high expressions of *KAR* and *KASII* genes improved the biosynthesis of stearic acid, which provided more precursors for the synthesis of carbon eighteen unsaturated fatty acids.
- The continuous high expression of *SAD* gene catalyses stearic acid desaturation to synthesize oleic acid and the continuous increased ratio of *SAD* to *FATB* gene expressions directly improved the fatty acid desaturation rate.
- The expression peaks of *FAD2*, *FAD3*, *FAD7* and *FAD8* genes simultaneously appeared in the periods of rapid biosynthesis of linoleic and linolenic acids and then the oleic acid was gradually desaturated to linoleic acid and linolenic acid.



## 3.4 Identification of key genes involved in the high accumulation of palmitoleic acid and oil in sea buckthorn berry pulp

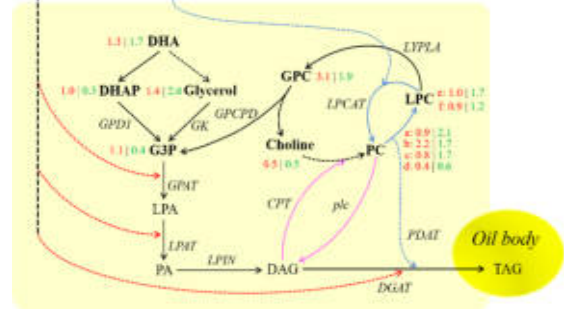
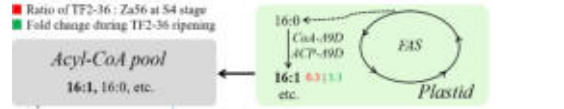
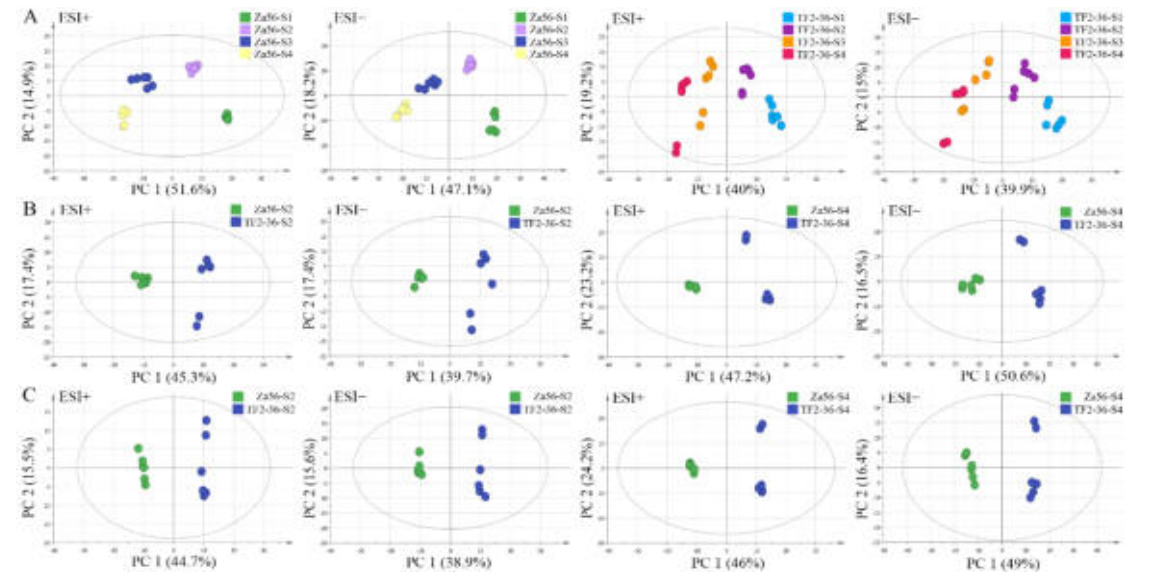
➤ The sustained high expression of *ACP-Δ9D* gene, which was related to oil, and the sustained low expression of *KASII* and *FAE1* genes, contributed to the enrichment of C16:1 fatty acids pulp.

➤ Non target metabolomics showed G3P, *GPD1* and *Δ9D* contributes to high oil content and high palmitoleic acid content of sea buckthorn pulp



Oil synthesis and fatty acid metabolism pathway in sea buckthorn pulp

Heat map of gene expression related to fatty acid formation and accumulation in berry pulp

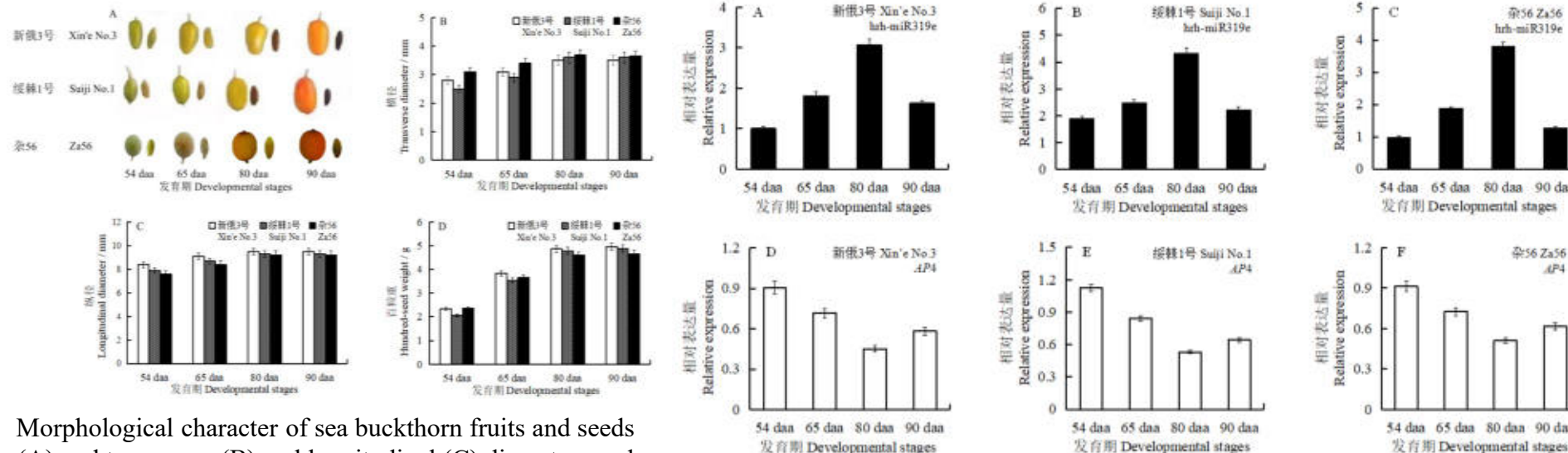


(Jian Ding, Chengjiang Ruan\*, Ying Guan, He Li, Wei Du, Shuang Lu, Xiufeng Wen, Ke Tang, Ye Chen. 2022. Nontargeted metabolomic and multigene expression analyses reveal the mechanism of oil biosynthesis in sea buckthorn berry pulp rich in palmitoleic acid. *Food Chemistry*, 374 : 131719)

(Jian Ding, Chengjiang Ruan\*, Wei Du, Ying Guan. RNA-seq data reveals a coordinated regulation mechanism of multigenes involved in the high accumulation of palmitoleic acid and oil in sea buckthorn berry pulp. *BMC Plant Biology*, 2019, 19: 207)

## 3.5 Sea buckthorn hrh-miR319e targeting *AP4* gene regulates seed development

- The results shows that *AP4*-3'UTR region and miR319e seed sequence were completely complementary. The vector of PCDNA3.1-hrh-miR319e+PGL0mir-*AP4* can significantly inhibit the luciferase activity ( $p < 0.001$ ).
- In developing seeds, the changes of hrh-miR319e expression level showed a trend of increasing at first and then decreasing, while the changes of *AP4* expression level first decreased and then increased. When hrh-miR319e expression reached its highest level at 80 daa (days after anthesis) ('Xin'e 3'—3.146, 'Suiji 1'—4.298 and 'Za 56'—3.892), the expression level of *AP4* had the lowest value ('Xin'e 3'—0.427, 'Suiji 1'—0.526 and 'Za 56'—0.451).
- Thus, sea buckthorn hrh-miR319e could target the 3'UTR region of *AP4*, there is negatively regulative relationship between them.

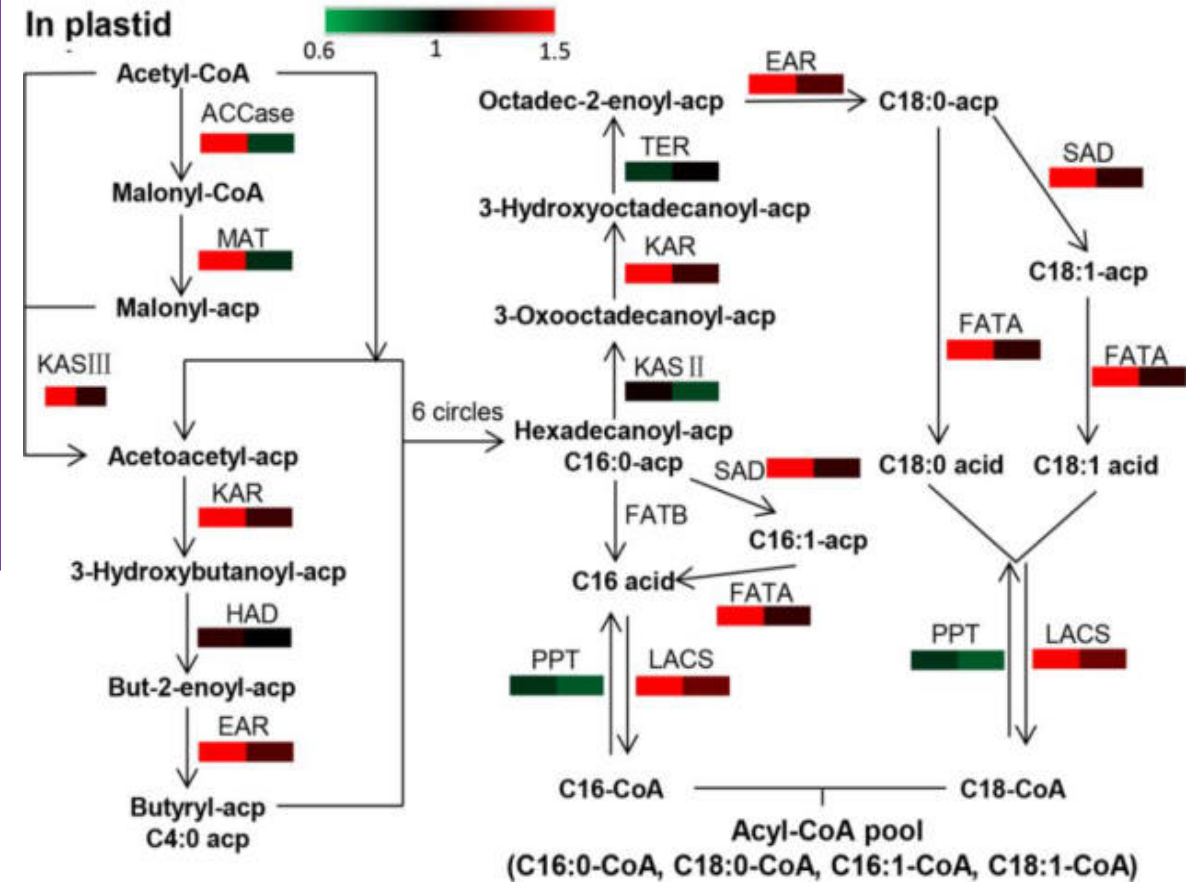
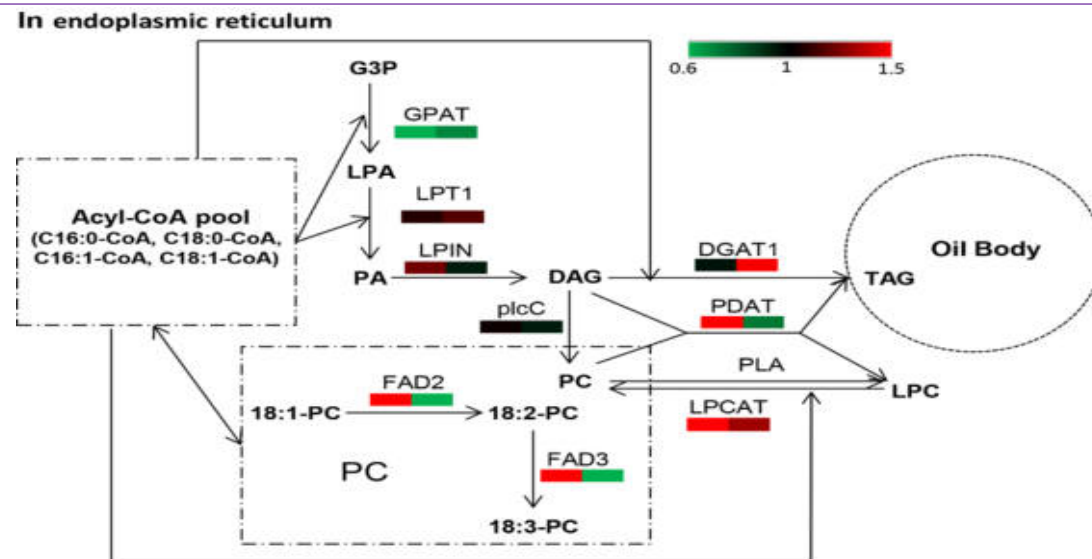


Morphological character of sea buckthorn fruits and seeds (A) and transverse (B) and longitudinal (C) diameters and hundred-seed weights (D) of seeds in different developmental stages

Changes of expression levels of hrh-miR319e and *AP4* of sea buckthorn seeds at different developmental stages

## 3.6 Proteomics identified several key enzymes for lipid metabolism in sea buckthorn

- Deep analysis results for the proteins identified and related pathways revealed initial fatty acid accumulation during whole-berry development.
- In the TAG metabolic pathway, **the effect of PDAT is more obvious in the early stage of fruit development**, which is conducive to increasing the proportion of unsaturated fat acids in fruit TAG, and **the DGAT is significantly increased in the late stage of development**, which is conducive to completing the transformation of fatty acids and intermediates to triglycerides.



Du Wei, Xiong Chao-Wei, Ding Jian, Nybom Hilde, Ruan Cheng-Jiang \*, Guo Hai. TMT-based Quantitative Proteomics of Developing Sea Buckthorn Berries Reveals Candidate Proteins Related to Lipid Metabolism. *Journal of Proteome Research*, 2019, 18(5): 1958-1969

## 3.7 Key flavonoid and lipid synthesis proteins in the sea buckthorn pulp

- In the flavonoid synthesis pathway, CHS and F3H were the main enzymes responsible for the difference in flavonoid content between the two cultivars.
- In the lipid synthesis pathway, LPIN, plcC and MGD were the main enzymes with different contents in the middle to late stages. Higher contents of LPIN and plcC in XE than in SJ could cause DAG to generate TAG from PC, since the difference in DGAT between the two cultivars was not significant.

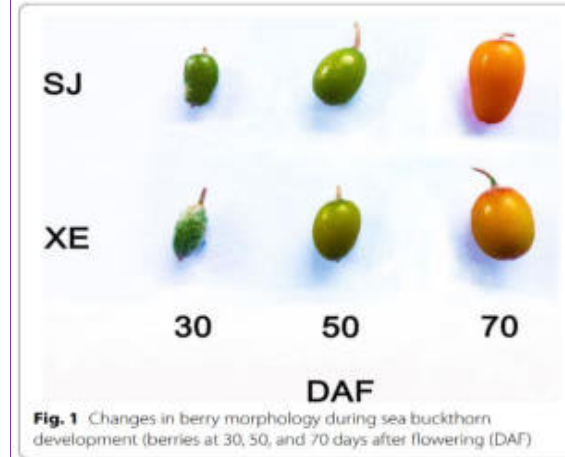


Fig. 1 Changes in berry morphology during sea buckthorn development (berries at 30, 50, and 70 days after flowering (DAF))

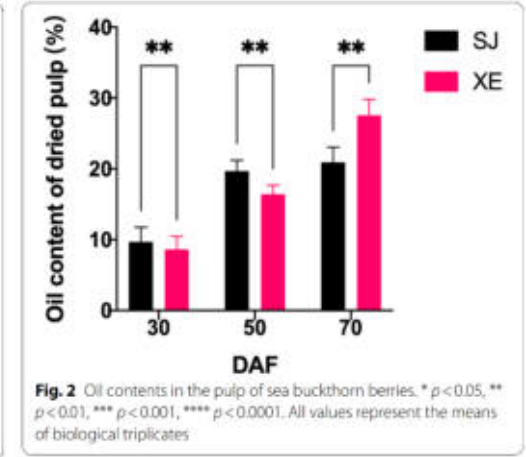


Fig. 2 Oil contents in the pulp of sea buckthorn berries. \*  $p < 0.05$ , \*\*  $p < 0.01$ , \*\*\*  $p < 0.001$ , \*\*\*\*  $p < 0.0001$ . All values represent the means of biological triplicates

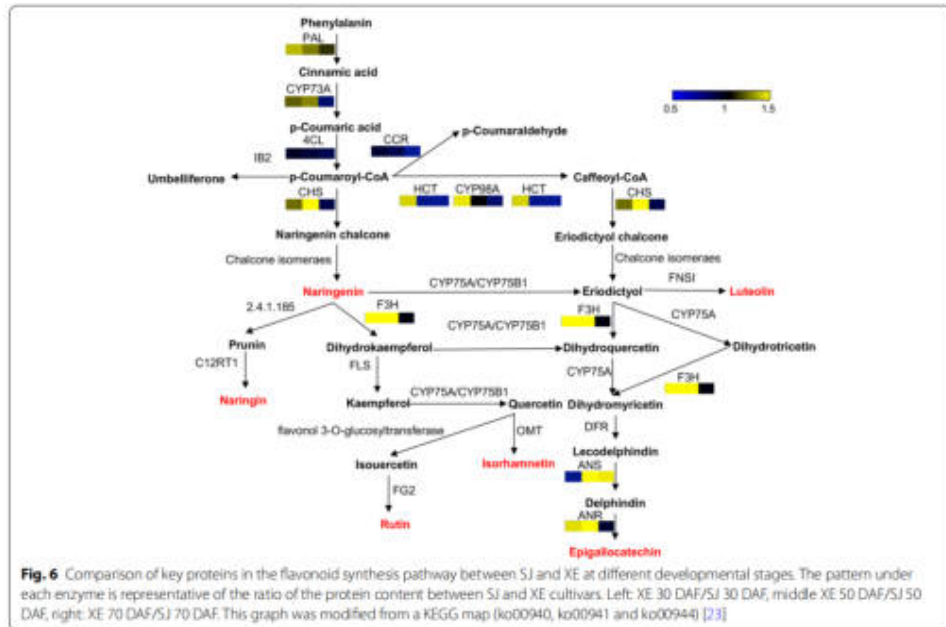


Fig. 6 Comparison of key proteins in the flavonoid synthesis pathway between SJ and XE at different developmental stages. The pattern under each enzyme is representative of the ratio of the protein content between SJ and XE cultivars. Left: XE 30 DAF/SJ 30 DAF, middle XE 50 DAF/SJ 50 DAF, right: XE 70 DAF/SJ 70 DAF. This graph was modified from a KEGG map (ko00940, ko00941 and ko00944) [23]

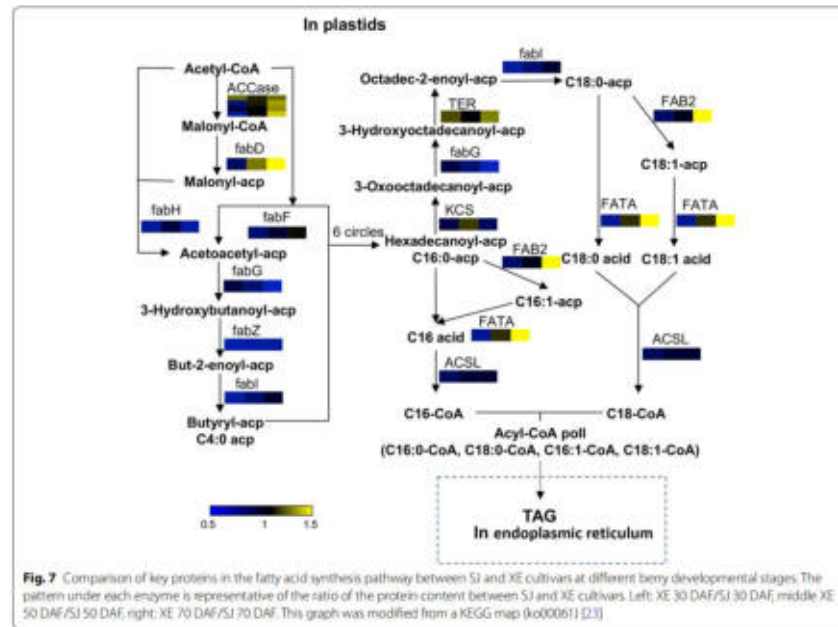
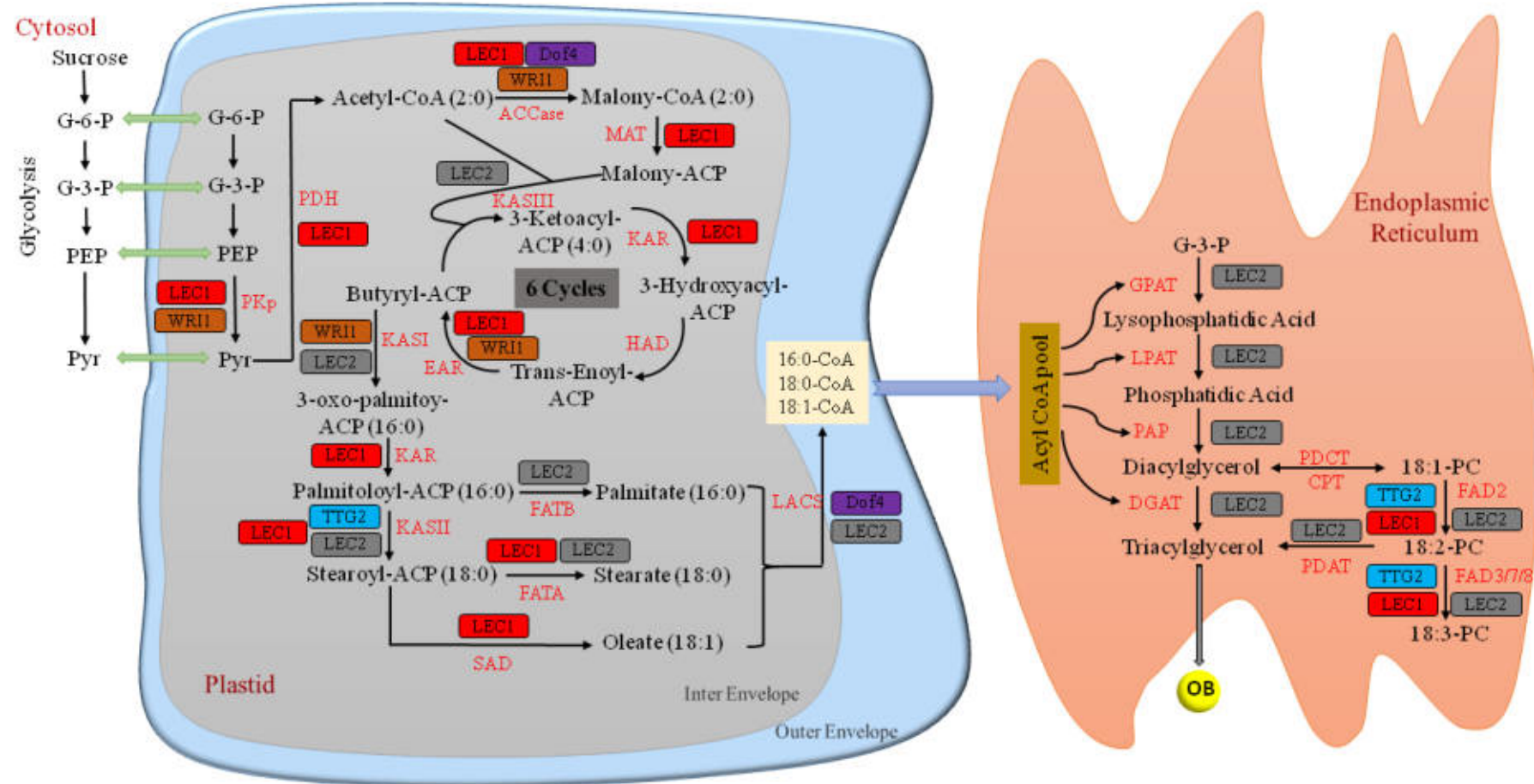


Fig. 7 Comparison of key proteins in the fatty acid synthesis pathway between SJ and XE cultivars at different berry developmental stages. The pattern under each enzyme is representative of the ratio of the protein content between SJ and XE cultivars. Left: XE 30 DAF/SJ 30 DAF, middle XE 50 DAF/SJ 50 DAF, right: XE 70 DAF/SJ 70 DAF. This graph was modified from a KEGG map (ko00061) [23]

Wei Du, Jian Ding, Shunguang Lu, Xiufeng Wen, Jianzhong Hu and **Chengjiang Ruan\***. Identification of the key favonoid and lipid synthesis proteins in the pulp of two sea buckthorn cultivars at different developmental stages. *BMC Plant Biology*, 2022, 22: 299

## 3.8 Identified transcription factor regulating key gene expression in lipid biosynthesis of sea buckthorn



Comparing transcriptomics and co-expression analysis showed main transcription factors involved in regulating lipid biosynthesis in sea buckthorn seeds, such as **ABI3, ABI4, AGL15, WRI1, FUS3, SWEET, Dof4, WRKY6, GRF8**, etc.





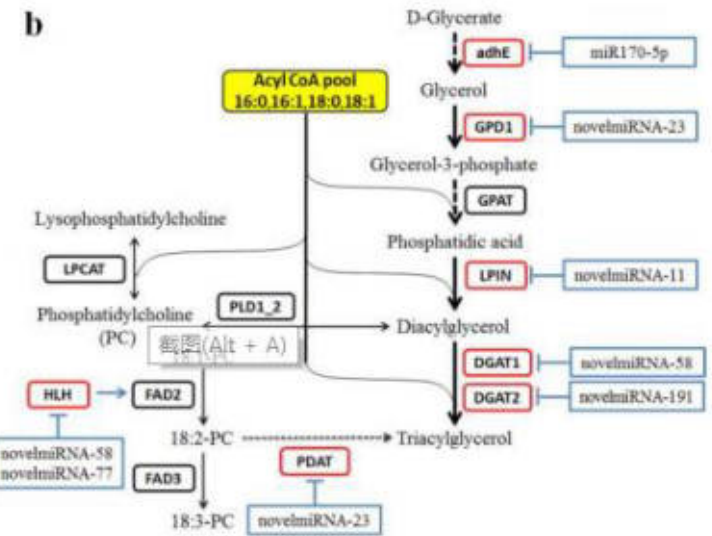
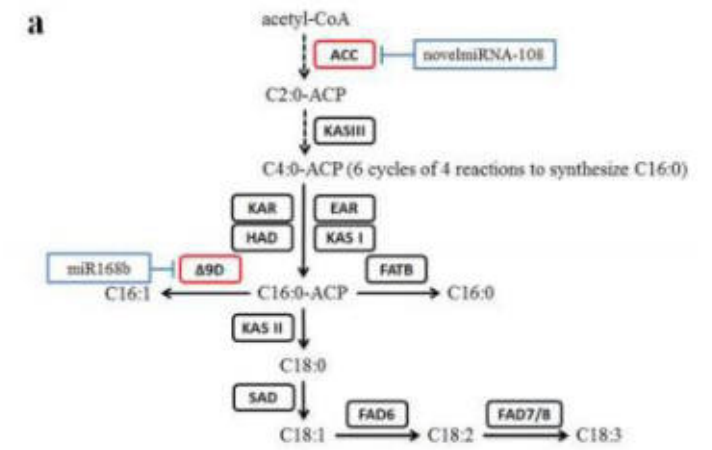
# 3. Formation mechanism of important traits in species of *Hippophae* genus



## 3.9 Identification of miRNA regulating expression of key genes involving in lipid biosynthesis in sea buckthorn

- Screened 19 (14 known and 5 new) miRNAs involved in regulating the accumulation of sea buckthorn oil synthesis, including miR164d-*ARF2*, miR168b-*Δ9D*, novelmiRNA-108-*ACC*, novelmiRNA-23-*GPD1*, novelmiRNA-58-*DGAT1* and novelmiRNA-191-*DGAT2*.

KEGG Pathway	miRNA name	Target ID	Annotation for targets	Gene name
Fatty acid biosynthesis	novelmiRNA-2	c141756_g2_12	long-chain acyl-CoA synthetase	ACSL
	novelmiRNA-108	c103701_g1_11	acetyl-CoA carboxylase carboxyl transferase	ACC
	novelmiRNA-110	c141756_g2_12	long-chain acyl-CoA synthetase	ACSL
Fatty acid elongation	novelmiRNA-170	c168561_g1_11	enoyl-CoA hydratase	ECHS
Fatty acid degradation	aly-miR170-5p	c60336_g1_11	alcohol dehydrogenase	adh
	novelmiRNA-2	c141756_g2_12	long-chain acyl-CoA synthetase	ACSL
	novelmiRNA-110	c141756_g2_12	long-chain acyl-CoA synthetase	ACSL
	novelmiRNA-170	c168561_g1_11	enoyl-CoA hydratase	ECHS
Biosynthesis of unsaturated fatty acids	gma-miR168b	c119361_g2_11	delta-9-desaturase	Δ9D
	novelmiRNA-58	c145891_g1_14	helix loop helix transcription factor	HLH
	novelmiRNA-77	c142283_g2_11	helix loop helix transcription factor	HLH
Glycerolipid metabolism	gma-miR164d	c154991_g1_11	dihydroxyacetone kinase	DAK
	novelmiRNA-11	c192054_g1_11	phosphatidate phosphatase	LPIN
	novelmiRNA-23	c133634_g3_15	phospholipid:diacylglycerol acyltransferase	PDAT
	novelmiRNA-58	c144982_g1_12	diacylglycerol O-acyltransferase 1	DGAT1
Glycero- phospholipid metabolism	novelmiRNA-191	c220405_g1_11	diacylglycerol O-acyltransferase 2	DGAT2
	novelmiRNA-11	c192054_g1_11	phosphatidate phosphatase	LPIN
	novelmiRNA-23	c138230_g1_13	glycerol-3-phosphate dehydrogenase	GPD1
Steroid biosynthesis	novelmiRNA-64	c176644_g1_11	lysophospholipid hydrolase	NTE
	novelmiRNA-10	c81203_g2_11	cycloartenol synthase	CAS
	novelmiRNA-58	c131674_g1_11	sterol-4-alpha-methyl oxidase	SMO2
	novelmiRNA-58	c144982_g1_13	sterol O-acyltransferase	SOAT
	novelmiRNA-179	c81203_g2_11	cycloartenol synthase	CAS
	novelmiRNA-224	c137936_g1_11	cycloartenol synthase	CAS
	novelmiRNA-224	c137936_g1_12	lanosterol synthase	LSS
Sphingolipid metabolism	novelmiRNA-232	c81203_g2_11	cycloartenol synthase	CAS
	zma-miR159b-3p	c262907_g1_11	beta-galactosidase	lacZ
	novelmiRNA-23	c130447_g1_13	neutral ceramidase	ASAH2
	novelmiRNA-108	c167130_g1_11	sphingolipid delta-4 desaturase	DEGS
	novelmiRNA-151	c209847_g1_11	beta-galactosidase	GLB1
novelmiRNA-170	c199679_g1_11	sphingomyelin phosphodiesterase 2	SMPD2	



(Jian Ding, Chengjiang Ruan\*, Ying Guan, Priti Krishna. Identification of microRNAs involved in lipid biosynthesis and seed size in developing sea buckthorn seeds using high-throughput sequencing. Scientific Reports, 2018, 8 : 4022)



## 3.11 LncRNAs involved in the oil synthesis of different organs of sea buckthorn

➤ The whole transcriptome association analysis found that four lncRNAs (LINC48098, LINC48300, LINC6093 and LINC9793) and two miRNAs (ath-miR172a and ath-miR858b) were involved in the regulation of 24 transcription factors (such as MYB, AP2/ERF and TCP family) that act on the formation of sea buckthorn fruit (Figure 2.23). Through transcription factor association analysis, a total of 5762 lncRNA miRNA TF modules were obtained, including LINC39069 ath miR157a-5p-SCL32, LINC35803 ath miR156i-BH094, LINC20980 ath miR398a-3p-IDM1, and LINC7158 novel\_186-ARF1, LINC21206-ath-miR858a-ERF6, LINC45543-novel\_The 226-WRI11 module regulates sea buckthorn fruit.

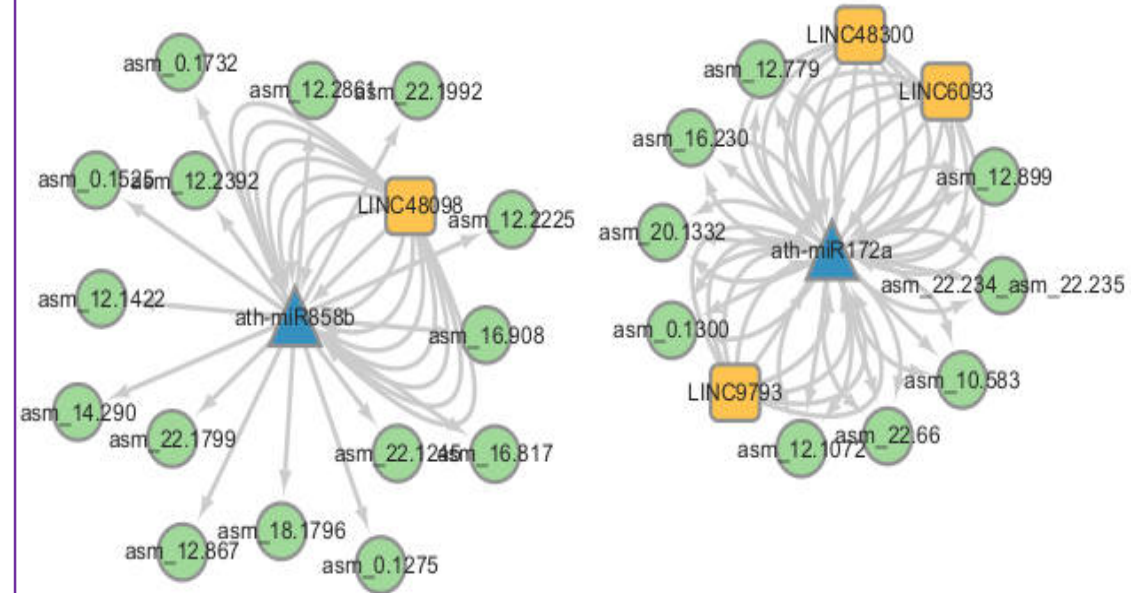


Figure 23 Regulation internet of lncRNA-miRNA-TF  
Circles represent transcription factors; Triangle represents miRNA; Rectangle represents lncRNA

43 regulatory modules formed among 41 lncRNAs, 8 miRNAs, and 6 target genes are related to lipid biosynthesis. The same lncRNA-miRNA module can target multiple genes, such as LINC31812-novel\_12 targets FATB, while LINC42952-ath-miR156i can also target FATB. Multiple functional modules related to oil synthesis and fatty acid formation and accumulation, such as LINC37656 targeting ath-miR396b-5p, LINC37732 targets ath-miR159c, LINC45223 targets ath-miR159c, Ath-miR396b-5p targeting *LACS6*, Ath-miR159c targets *FAD3*, etc.



## 4. Genetic improvement strategies of species in *Hippophae* genus



Improvement traits	Comparison species/subspecies
Big fruit, big seed, high grain weight	<i>H. tibetana</i> /ssp. <i>mongolica</i> vs. ssp. <i>sinensis</i>
High oil content in seeds	<i>H. gyantsensis</i> / <i>H. tibetana</i> / <i>H. neurocarpa</i> vs. ssp. <i>sinensis</i>
High palmitoleic acid and low palmitic acid	ssp. <i>sinensis</i> vs. <i>H. gyantsensis</i> //
Tolerance to water logging	ssp. <i>sinensis</i> South of China vs. North of China; ssp. <i>yunnanensis</i> / <i>H. tibetana</i> / <i>H. neurocarpa</i> / <i>H. salicifolia</i> vs. ssp. <i>mongolica</i>
Tolerance to weather with high temperature and humid	ssp. <i>mongolica</i> /ssp. <i>sinensis</i> vs. ssp. <i>yunnanensis</i> / <i>H. tibetana</i> / <i>H. neurocarpa</i> / <i>H. salicifolia</i>
Adaptation to high altitude	<i>H. tibetana</i>
Tolerance to drought	Different ecotypes of ssp. <i>turkestanica</i>

### Future modern techniques for sea buckthorn breeding:

crossing, genetic transformation, gene editing.....



## 5. Several interesting questions



- ① It is necessary to construct a super-pangenome of all species and subspecies in *Hippophe* genus ;
- ② How are species formed? Reproductive isolation, geographical isolation?
- ③ How hybridization forms new species ?
- ④ How to adapt to high altitude ?
- ⑤ How to successfully transplant high-altitude species to low altitude habitats ?
- ⑥ How does climate change affect the population changes of seabuckthorn?



# Welcome you visit my Lab.



**Prof. Dr.  
Chengjiang  
Ruan**



**Associate  
Prof. Dr.  
He Li**



**Associate  
Prof. Dr.  
Jian Ding**



**Associate  
Prof. Dr.  
Jingbin Li**



**Associate  
Prof. Dr.  
Yuhui Hong**



**Engineering  
Ms. Wei Du**

## Thank you for your attention!

